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## Relationship between the antibacterial and immunological activities of houttuynonate homologues and their surface activities

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Based on the microbial model (*Staphylococcus aureus* and *Bacillus subtilis*) and the experiment of mouse carbon granular clearance, the relationship between antibacterial and immunological activities of houttuynonate (HOU) homologues and their surface activities were studied. The results showed that with elongating the carbon chain of HOU homologues, the surface activities and bacteriostasis and immunological activities would be increased. It is suggested that the pharmacological effect of HOU homologues could be realized by the hydrophobic interaction between HOU homologues and membrane proteins of bacteria and cell.

**Keywords:** Houttuynonate (HOU) homologues; Surface tension; Bacteriostasis; Immune activity

### 1. Introduction

*Houttuynina cordata Thunb* is used as a traditional Chinese herbal medicine, which can improve body immunity, clear away heat, eliminate carbuncles and purulence, and diminish inflammation [1]. Decanoyl acetaldehyde is a main antibacterial component in *Houttuynina cordata Thunb*, named houttuynine. However, the content of houttuynine in *Houttuynia cordata Thunb* is minimal, and its water-extracting rate is about 0.00049% [2]. In addition, houttuynine is insoluble in water, and unstable *in vitro* [3]. Therefore, houttuynine has been transformed into decanoyl acetal sodium sulfite, named houttuyninum, which has better water solubility and stability, and has antibacterial activity as good as houttuynine [4]. Subsequently, houttuyninum has been substituted for houttuynine in the clinical setting.

According to the molecular structure of houttuyninum, dodecanoyl acetal sodium sulfite has been synthesized and named sodium new houttuynonate [5]. At present, houttuyninum

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and sodium new houttuynonate are used to treat respiratory system diseases, influenza, acute or chronic bronchitis and pneumonia, etc. [3,6].

In our laboratory, four kinds of houttuynonate homologues ( $C_nH_{2n+1}C(O)CH_2C(OH)SO_3Na$ ,  $n = 6,8,10,12$ ; abbreviated as HOU- $C_n$ ) were synthesized by changing the length of alkyl chain based on the structure of houttuyninum (HOU- $C_{10}$ ) [7].

HOU- $C_{10}$  is a typical surfactant, which has a hydrophobic alkyl group and a hydrophilic vitriol group. Up to now, there is no report on the relationship between the antibacterial and immune activities of HOU- $C_n$  and their surface activities. In our experiment, *Staphylococcus aureus* and *Bacillus subtilis* were used as the microbe model to study the antibacterial activity of HOU- $C_n$ , and white mice were used as the animal model to investigate the effect of carbon chain length on immunological function. By investigating the relationship between the surface activity and biological activity of HOU- $C_n$ , more potent and lower side-effect medicines might be obtained.

## 2. Results and discussion

### 2.1 Surface tension of HOU- $C_n$

According to their structure, HOU- $C_n$  belong to surfactants. The surface tension of HOU- $C_n$  versus logarithm of their concentration is shown in figure 1. According to the turning point of curves, critical micelle concentration (CMC) of HOU- $C_n$  can be determined. Therefore, the CMCs of HOU- $C_6$ , HOU- $C_8$ , HOU- $C_{10}$  and HOU- $C_{12}$  were  $5.32 \times 10^{-3}$  mol/L,  $1.41 \times 10^{-3}$  mol/L,  $3.88 \times 10^{-4}$  mol/L and  $1.06 \times 10^{-4}$  mol/L, respectively.

Figure 2 shows that the carbon number of HOU- $C_n$  versus logarithm of their CMC is a linear relationship. Therefore, the free energy of HOU- $C_n$  forming micelle can be calculated based on its CMC, according to the formula as follows [8]:

$$\Delta G = 2.303 \times RT \times \log(CMC)$$

The longer the carbon chain of HOU- $C_n$  is, the smaller are the CMC and the  $\Delta G$ . Correspondingly, the easier the micelle that would be formed and the stronger the hydrophobic

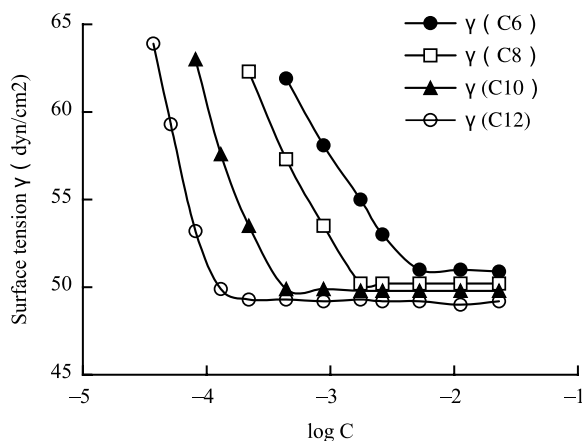


Figure 1. Surface tension versus logarithm of concentration of HOU- $C_n$ .

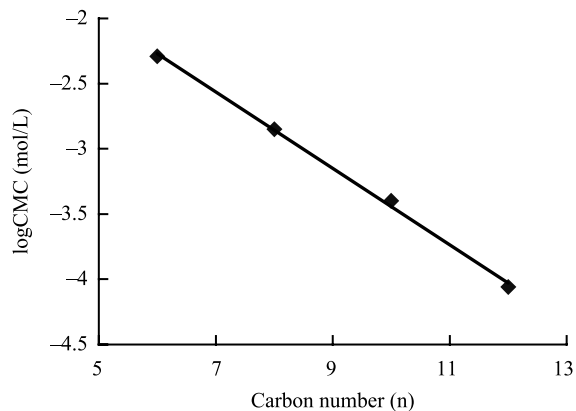


Figure 2. Relationship between carbon number of HOU- $C_n$  and its logCMC

interaction that would exist with other hydrophobic groups. It was indicated that the hydrophobic ability increased along with elongation of the carbon chain.

## 2.2 Bacteriostasis of HOU- $C_n$ to *Staphylococcus aureus*

The lower the absorbency of the media is, the more potent is the bacteriostatic effect of HOU- $C_n$ . From figure 3, the bacteriostasis of HOU- $C_n$  to *Staphylococcus aureus* would obviously increase along with the increase of HOU- $C_n$  concentration, and the bacteriostatic effect is directly proportional to the carbon number of HOU- $C_n$ . The longer the carbon chain of HOU- $C_n$  is, the stronger are their bacteriostasis to *Staphylococcus aureus* and the lower are the concentrations of fully inhibiting *Staphylococcus aureus*. The full-inhibition concentration of HOU- $C_6$ , HOU- $C_8$ , HOU- $C_{10}$  and HOU- $C_{12}$  was 0.553, 0.363, 0.226 and 0.153 mg/ml, respectively (figure 3).

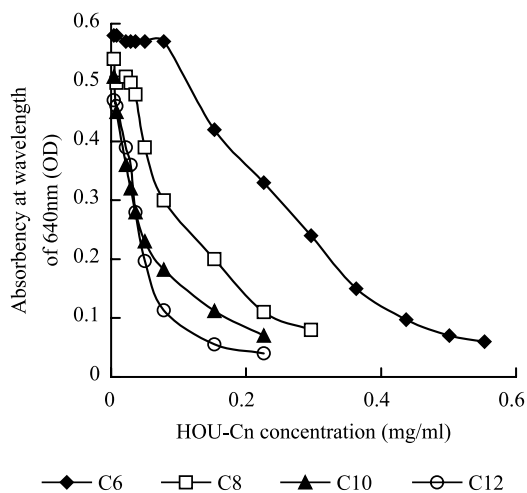


Figure 3. Bacteriostasis of HOU- $C_n$  to *Staphylococcus aureus*.

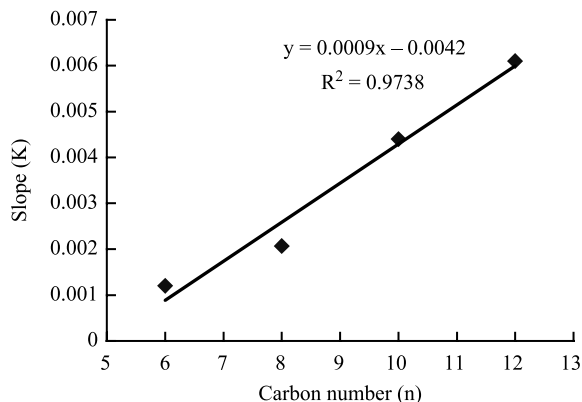


Figure 4. Carbon number of HOU-C<sub>n</sub> versus bacteriostatic activity to *Staphylococcus aureus*.

From figure 4, the slope of precipitous curves versus carbon number of HOU-C<sub>n</sub> indicated that the slope of curve would be increased in parallel with the increase of carbon number of HOU-C<sub>n</sub>. This indicates that bacteriostasis of HOU-C<sub>n</sub> to *Staphylococcus aureus* was positively proportional to the carbon number of HOU-C<sub>n</sub>. In these four kinds of HOU-C<sub>n</sub>, the bacteriostatic activity of HOU-C<sub>12</sub> was the strongest.

### 2.3 Bacteriostasis of HOU-C<sub>n</sub> to *Bacillus subtilis*

Figure 5 shows the bacteriostasis of HOU-C<sub>n</sub> to *Bacillus subtilis*. The bacteriostasis to *Bacillus subtilis* was similar to that to *Staphylococcus aureus* (shown in figure 3). Within a certain concentration, bacteriostasis of HOU-C<sub>n</sub> would be strengthened along with the concentration increase, and bacteriostasis of HOU-C<sub>n</sub> would also be strengthened alongside elongation of the carbon chain.

Figure 6 shows that the relationship between the slope of the precipitous curve was directly proportional to the carbon number of HOU-C<sub>n</sub>. It indicates that the longer the carbon number

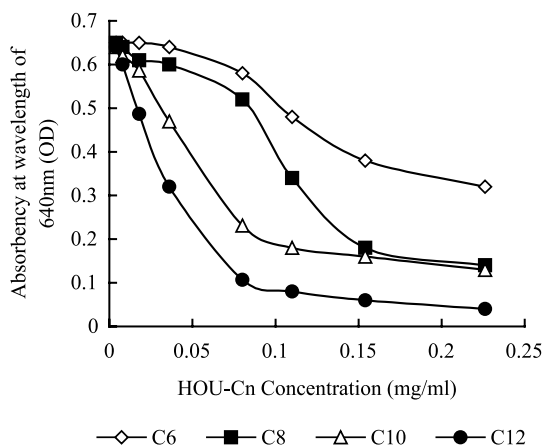


Figure 5. Bacteriostasis of HOU-C<sub>n</sub> to *Bacillus subtilis*.

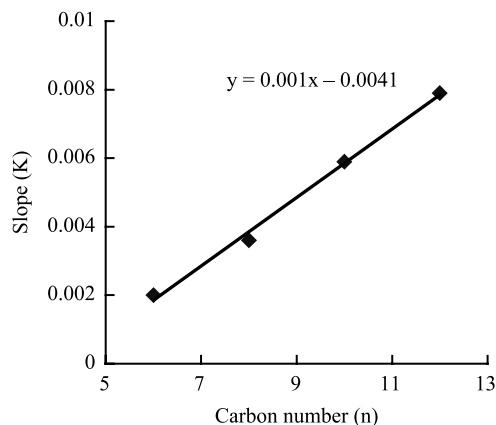


Figure 6. Carbon number of HOU- $C_n$  versus bacteriostatic activity to *Bacillus subtilis*.

of HOU- $C_n$  is, the more potent is the bacteriostasis of HOU- $C_n$  to *Bacillus subtilis*. Similarly, the bacteriostatic activity of HOU- $C_{12}$  was the highest.

These results showed that hydrophobic ability and bacteriostasis of HOU- $C_n$  would be increased along with an elongating carbon chain, and that bacteriostasis of HOU- $C_n$  to *Staphylococcus aureus* and *Bacillus subtilis* was positively proportional to their hydrophobic ability.

#### 2.4 Determination of immune activity of HOU- $C_n$

Table 1 shows that after injecting 3 mg/kg of HOU- $C_n$  into mice for 7 days, HOU- $C_n$  could increase the carbon granular clearance rate  $K$  of macrophage, especially HOU- $C_{12}$ , and that the phagocytic function of macrophages in mice injected with HOU- $C_n$  was enhanced. There was little influence of HOU- $C_n$  on thymus gland index and spleen index, suggesting that HOU- $C_n$  could improve the immunological function of mice.

The effects of different concentrations of HOU- $C_n$  on immune ability are shown in figure 7. Under the same dose, immune activity of mice rose steadily alongside the elongation of the carbon chain. In the dose range of 1.5–6 mg/kg of HOU- $C_n$ , immune ability increased along with the rise of the dose. The results differed from literature reports, which found that when the dose was higher than 6 mg/kg, the immune function of mice was enhanced along with a decrease of the dose of HOU- $C_{10}$  [6].

Table 1. Effects of sodium houttuynonate homologues on the immune organs of mice ( $\bar{x} \pm s$ ).

Group	Carbon granular clearance rate $K$	Thymus gland index (mg/g)	Spleen index (mg/g)
CK	0.0129 $\pm$ 0.0031	1.3997 $\pm$ 0.1223	3.9762 $\pm$ 0.1513
$C_6$	0.0197 $\pm$ 0.0033*	1.3818 $\pm$ 0.1369	4.0175 $\pm$ 0.1272
$C_8$	0.0241 $\pm$ 0.0038*	1.4046 $\pm$ 0.1121	4.0322 $\pm$ 0.1315
$C_{10}$	0.0286 $\pm$ 0.0037*	1.3802 $\pm$ 0.1215	4.0279 $\pm$ 0.1411
$C_{12}$	0.0330 $\pm$ 0.0155**	1.4051 $\pm$ 0.1198	4.0772 $\pm$ 0.1331

Dose of HOU- $C_n$  is 3 mg/kg. \* $p < 0.05$ , \*\* $p < 0.01$  compared with CK.

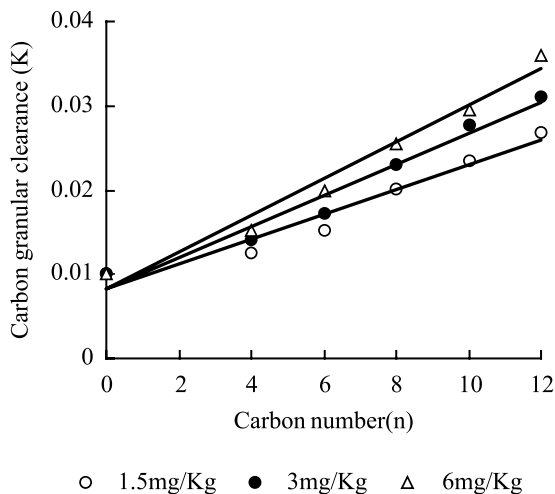


Figure 7. Carbon number versus immune activity of HOU-C<sub>n</sub>.

Our results showed that improved immunity of HOU-C<sub>n</sub> was positively proportional to the length of the carbon chain, and that the biological activity of HOU-C<sub>n</sub> was directly proportional to its hydrophobic ability.

## 2.5 Discussion

The results of microbial and animal experiments showed that the bacteriostasis and immune function of HOU-C<sub>n</sub> were proportional to the length of their carbon chain. A surface tension test indicated that the hydrophobic activity of HOU-C<sub>n</sub> was directly proportional to their carbon chain length.

Previous studies [8] showed that a surfactant would bind with the hydrophobic site of protein or enzyme, causing the conformation and biological activity of the protein to be changed. Further evidence reported [9] that the longer the carbon chain of the surfactant, the stronger was the ability of their denaturalization to enzyme and protein. HOU-C<sub>n</sub> was a typical surfactant, whose bacteriostasis and immune activities were positively proportional to their hydrophobic activity, which was similar to that of surfactant and enzyme [8]. This suggested that the pharmacological effect of HOU-C<sub>n</sub> could be caused by the hydrophobic combination between HOU-C<sub>n</sub> and the membrane protein of bacteria and cell.

The stronger the hydrophobic activity of HOU-C<sub>n</sub> is, the better the immune and antibacterial activity of HOU-C<sub>n</sub> are. It is speculated that HOU-C<sub>n</sub> could combine with the hydrophobic site of membrane protein by hydrophobic interaction, because the stronger hydrophobic activity of HOU-C<sub>n</sub> would be more beneficial if combined with bacteria and cell, further improving physiological activity.

Experimental results showed that in the dose range of 1.5–6 mg/kg of HOU-C<sub>n</sub>, immune ability was increased alongside the rise of the dose. But when the dose was too high, the immune function of mice decreased in parallel with the increase of the dose of HOU-C<sub>10</sub> [6]. Within a range of a certain concentration, a surfactant would make the conformation of

protein more flexible and increase enzyme activity. However, when its concentration was too high, it would make the conformation of protein change to excess and further cause enzyme activity to be lost.

### 3. Experimental

#### 3.1 Synthesis of sodium houttuyfonate homologues

Sodium houttuyfonate homologues ( $C_nH_{2n+1}C(O)CH_2C(OH)SO_3Na$ , abbreviated as HOU- $C_n$ ) were synthesized according to Li *et al.* [7], including hexanoyl acetal sodium sulfite (HOU- $C_6$ ), octanoyl acetal sodium sulfite (HOU- $C_8$ ), decanoyl acetal sodium sulfite (HOU- $C_{10}$ ) and dodecanoyl acetal sodium sulfite (HOU- $C_{12}$ ). All samples in the experiments were recrystallised five times, and their contents were more than 97% analysed according to the standard methods of the Chinese Pharmacopoeia.

#### 3.2 Determination of surface tension [9]

The concentrations of HOU- $C_n$  were controlled at 0.025 mol/L. HOU- $C_n$  decompose in water and release sodium hydrogen sulphite. When measuring their surface tension, 0.5 mol/L of sodium hydrogen sulphite should be added into the above solutions to keep these compounds stable. The water solubility of HOU- $C_{10}$  and HOU- $C_{12}$  was very low, therefore the temperature of the solution should be controlled at  $55 \pm 0.5^\circ\text{C}$  when measuring their surface tension.

#### 3.3 Bacteriostasis of HOU- $C_n$ [10]

*Staphylococcus aureus* and *Bacillus subtilis* were grown on 5 ml of liquid media (pH 7.2) for about 6 h each, and adjusted to  $10^5$ – $10^6$  bacteria per milliliter under the microscope. HOU- $C_n$  was diluted into different concentration solutions with liquid media. Liquid bacteria incubated for 6 h were planted into these HOU- $C_n$  media. After being incubated for another 24 h, the absorbency of the above media was determined at 640 nm by a spectrophotometer, then curves of HOU- $C_n$  versus bacteriostasis were calculated in which HOU- $C_n$  concentration was the abscissa and absorbency was the ordinate. The beginning bacteriostatic concentration was taken as that one just as absorbency of the media decreased, and the full inhibition concentration when absorbency of the media was equal to that of the control. Fifty microlitres of HOU- $C_n$  media were taken from the test tube in which bacteria had been fully inhibited and planted in the plate media, then incubated for about 24 h. The bactericidal concentration was determined if there had been no bacteria in the plate media.

#### 3.4 Immune activity of HOU- $C_n$

Healthy mice (18–22 g) were randomly divided into 13 groups, each containing 10 mice. Therapy groups included HOU- $C_n$ . Every kind of HOU- $C_n$  was divided into three gradient groups, according to concentrations of 0.1, 0.2 or 0.4 mg/ml injections of HOU- $C_n$ .



In therapy groups, every mouse was injected with 0.3 ml of HOU-C<sub>n</sub> and the control group was injected with the same dose of physiological saline. In fact, the dose of HOU-C<sub>n</sub> injected into mice every time was 1.5, 3 or 6 mg/kg. Every mouse was injected for 7 days. The day after the last injection, every mouse was weighed, and then injected in the tail vein with diluted India ink at a dose of 0.1 ml/10 g-body weight. Blood (20 µl) was taken with a capillary at 3 and 10 min after injection respectively, and dissolved in 1.5 mL of 0.1% Na<sub>2</sub>CO<sub>3</sub> and shaken up evenly. Absorbency of the above solution was measured at the wavelength of 680 nm.

After taking blood, the mice were killed humanely and the liver, spleen and thymus gland taken out and weighed. Immune ability was determined according to the carbon granular clearance rate (*K*) [6], revised index, thymus gland index and spleen index.

The Carbon granular clearance rate *K* was calculated by following equation:

$$K = \frac{\log A_1 - \log A_2}{t_2 - t_1}$$

### 3.5 Statistical analysis

Experimental data were analysed by DPS statistical software.

### References

- [1] S. Li, Q.H. Yu, Z.Y. Chu. *Chin. Pharm. Bull.*, **14**, 442 (1998).
- [2] X.Y. Hao, L. Li, Z.H. Ding. *Acta Bot. Yunnan.*, **17**, 350 (1995).
- [3] S.C. Wei, L.J. Xu, Q. Zeng. *Chin. Pharm. J.*, **34**, 167 (1999).
- [4] L. Shao, Q.H. Yu, Q. Huang, S.H.S. Liu. *Chin. Pharm. Bull.*, **17**, 51 (2001).
- [5] C.H.Q. Yuan. *Exploitation and Utilization of Natural Medicine Resource*, pp. 1569–1575, Jiangsu Science and Technique Public House, China (2000).
- [6] A.F. Du, S.H. Hu, W.H. Bao. *J. Trad. Chin. Vet. Med.*, **17**, 7 (1998).
- [7] X.G. Li, L.J. Yuan, H.M. He. Advanced water-soluble houttuynine. *China invention patent application* No. 2004100044932 (2004).
- [8] X.G. Li. *Acta Biophys. Sin.*, **11**, 13 (1995).
- [9] X.G. Li. *Physical Chemistry of Surfactants and their Application on Agriculture*, pp. 52–70, Southwest China Normal University Public House, China (1997).
- [10] H. Li, H.C. Jiang, G.L. Zhou. *Acta Botanica Yunnanica*, **24**, 95 (2002).